

Curcumin as an Environmental Potent Antioxidant Decreases Risk of Artherosclerosis

Parastoo Lourestanpour^a, Hossein Babaahmadi-Rezaei^{b*}, Kahin Shahanipour^a

^aDepartment of Biochemistry, Falavarjan Branch, Islamic Azad University, Esfahan, Iran.

^bCellular and Molecular Research Center, Department of Clinical Biochemistry, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.

*Correspondence should be addressed to Mr. Hossein Babaahmadi Rezaei, Email: hbabaahmadi@gmail.com

A-R-T-I-C-L-E-I-N-F-O

Article Notes:

Received: Aug. 20, 2016

Received in revised form:
Nov. 20, 2016

Accepted: Dec. 19, 2016

Available Online: Jan 1,
2017

Keywords:

Bovine aortic endothelial cells
Curcumin
H₂O₂
PDGF
Oxidative stress
Iran.

A-B-S-T-R-A-C-T

Background & Aims of the Study: Oxidative stress increases platelet-derived growth factor (PDGF) gene expression in endothelial cells that contributes to vascular dysfunction and atherosclerosis. Oxidative stress generates by dys-regulated redox balance between ROS producing systems and antioxidant systems. Also, Curcumin (Cur) as a main part of turmeric has anti-inflammatory, antioxidant, anticancer and antitumor effects. This study was conducted to test the Curcumin as an environmental potent antioxidant decreases risk of artherosclerosis.

Materials and Methods: This experimental study was conducted during 2015 in Iran. Cultured bovine aortic endothelial cells were incubated with hydrogen peroxide (H₂O₂) (20, 40 and 80 μM) and Curcumin (10 μM) for 24h. Then, the level of PDGF gene expression was analyzed by Real-Time PCR in untreated and treated cells.

Results: The results demonstrated significant increase in the level of PDGF gene expression in H₂O₂ treated groups versus control. Also, treated groups with H₂O₂-Curcumin showed notable decrease in the level of PDGF gene expression compared with H₂O₂ treated groups.

Conclusion: Our results support valuable data about the application of Curcumin for protection against atherosclerosis.

Please cite this article as: Lourestanpour P, Babaahmadi-Rezaei H, Shahanipour K. Curcumin as An Environmental Potent Antioxidant Decreases Risk of Artherosclerosis. Arch Hyg Sci 2017;6(1):105-10.

Background

Atherosclerosis or plaque build-up in the arteries as one of the serious health problems causes by various conditions such as high blood pressure and high level of cholesterol (1). Recent studies demonstrated remarkable data about the role of oxidative stress in atherosclerosis (1,2). There is an increase on the level of ROS production during cardiovascular diseases (2). The initial step of atherosclerosis is production of free radicals which can induce endothelial cells dysfunction. Oxidation of LDL is the most important cause

of oxidative stress. Two most sources of ROS are smooth muscle cells and macrophages (2,3). This fact suggests powerful ability of herbal antioxidant for reducing oxidative stress and risk of oxidative stress-induced atherosclerosis in capable person. Turmeric rhizome has yellow color and uses widely as a spice and for treating different pathogenic conditions such as body pain, intestinal worms, diarrhea, intermittent, fevers, hepatic disorders, biliousness, urinary discharges, dyspepsia, inflammations, constipation, leucoderma and amenorrhea (4-6). Furthermore, the potent effects of turmeric rhizome are related to its

components with strong anti-oxidant activities including Curcumin (7,8).

The levels of enzymatic anti-oxidant improve following Curcumin treatment in animals undergoing irradiation. Curcumin treatment before irradiation shows a significant increase in glutathione concentration and activities of both the glutathione peroxidase and superoxide dismutase in the irradiated mouse skin. Also, lipid peroxidation as a main mark of oxidative stress decreased in the skin of these animals (8). There is an evidence that shows Curcumin acts as potent free radicals scavenger in vivo in the brain. This antioxidant has neuroprotective effects against oxidative stress. Curcumin can inhibit parkinsonian neurotoxin –induced neuronal death in brain by anti-oxidant activity (9). In addition, Curcumin reduces DNA fragmentation, enzymes activities and lipid peroxidation in liver of lambda cyhalothrin (LCT)-intoxicated rats. Prevent DNA fragmentation, protect against oxidative stress and scavenge free radicals are the main results of Curcumin (10). Recently, based on the result of different study, Curcumin has significant effects on high fat diet and high cholesterol (4,5). Dietary Curcumin modulates high fat diet-induced atherosclerosis, steatohepatosis (4) and expression of genes involved in leukocyte adhesion and trans-endothelial migration in mice (5).

Aims of the study:

Our aim was determination of inhibitory effects of anti-oxidant on PDGF-mediated atherosclerosis in the endothelial cells which exposed to oxidative stress.

Materials & Methods

This experimental study was conducted during 2015 in Iran.

Cell culture

Cell lines of bovine aortic endothelial cells (BAECs) were cultured in DMEM (Gibco, Germany) containing FBS (Gibco, Germany) and penicillin/streptomycin (Gibco, Germany).

Incubation and Treatment

Suspension (containing 12×10^4 cells) was incubated for 24-48 h in CO₂ incubator (Jeltaghiz, Iran). Then, culture media of each well was exchanged with fresh culture media containing FBS and plate was incubated for 24-48h.

Also, all sample preparation was performed in a laminar flow hood (class II, Jeltaghiz, Iran). Cell sample was treated with Curcumin and H₂O₂ (Merck, Germany).

Cell collection

Cells were collected from culture after incubation (Incubator, NAPCO, USA). So, whole culture media was collected from surface of plate wells. Then, trypsin (Idehdist, Iran) was added to wells and plate was incubated. Culture media (Gibco, Germany) was added to wells while the component of wall was centrifuged (ROTOFIX32A, Hettich, Germany). Then, plate was used for RNA extraction and Real-Time PCR.

RNA extraction and Real-Time PCR

The aim of this method was the study of PGDF- β mRNA expression as a main marker for atherosclerosis. Total RNA was extracted from samples by RNeasy plus mini (QIAGEN, China), according to manufacturer's catalog.

RNA used for generated cDNA, using the Prime Script 1st strand cDNA Synthesis Kit (Takara, Japan) and amplified by real-time PCR, using a SYBR green Real time PCR (Takara, Japan) and the ABI Step One Plus real-time PCR instrument (Applied Biosystems, USA). Primers list are shown in the Table 1. Primers were designed by Allel ID version 6. Control mixture consisted of PCR mixture without cDNA.

Table 1) Primer sequences used for Real-Time PCR (5' to 3').

Primers	Genes
F: CAGAGCCAGCCGTGATT	PDGF
R: GCCACACCAGGGAAGTTAGC	
F: GGACACGGACAGGATTGACAG	18S
R: ATCGTCCACCACTAAGAACG	

Statistical analysis

The results of Real-Time PCR were analyzed by Delta CT formula and Rest version 2009.

The statistical analyses were performed, using the SPSS 20 (SPSS Inc., Chicago, IL, USA). One-way ANOVA (Tukey Multiple comparison) was used to test differences between various means (post hoc analysis LSD test). All experimental data were presented as the mean±SD. The level of significance for all tests was set at P<0.05.

Results

Real-Time PCR was used to evaluate PDGF-β gene expression in treated and untreated Bovine aortic endothelial cells. Comparative gene expression profiling analysis is useful in discovering differentially expressed genes associated with various agents and diseases (11). The levels of PDGF-β mRNA expression were different in experimental groups. The results showed in Table 2.

There were 6 groups in this research as; G1 (Control): untreated group, G2 (Group 2): treated with 20 μM H₂O₂ (for 24h), G3 (Group

3): treated with 40 μM H₂O₂ (for 24h), G4 (Group 4): treated with 20 μM H₂O₂ and 10 μM Curcumin (for 24h), G5 (Group 5): treated with μM H₂O₂ and 10 μM Curcumin (for 24h) and G6 (Group 6): treated with 80 μM H₂O₂ and 10 μM Curcumin (for 24h).

The results demonstrated a significant increase in the level of PDGF-β mRNA expression in group 2 (20 μl H₂O₂) compared with control and group 4 (20 μM H₂O₂-10 μM Curcumin) (p<0.05) (Figure 1). But, group 3 (40 μM H₂O₂) didn't show a significant increase versus control.

In addition, 40 μM H₂O₂-10 μM Curcumin treated group (group 5) demonstrated a decrease in PDGF-β mRNA expression compared with 40 μM H₂O₂ treated group (group 3) (Figure 2).

PDGF-β mRNA expression increased in response to 80 μM H₂O₂-10 μM Curcumin (group 6) versus groups 3, 5 and control (Figure 2). But, this change was not significant versus other groups. Unexpectedly, the results of groups 2 and 6 were close.

Table 2) The results of Real-Time PCR in control and treated groups

Groups	Group 1	Group 2	Group 3	Group 4	Group 5	Group 6
PDGF-B mRNA expression	1 ^a	3.74±0.37 ^b	1.82±1.15 ^{a,b}	0.72±0.59 ^a	1.64±0.32 ^{a,b}	3.28±0.64 ^{a,b}

Significant differences at p < 0.05 are shown by different letters in each column.

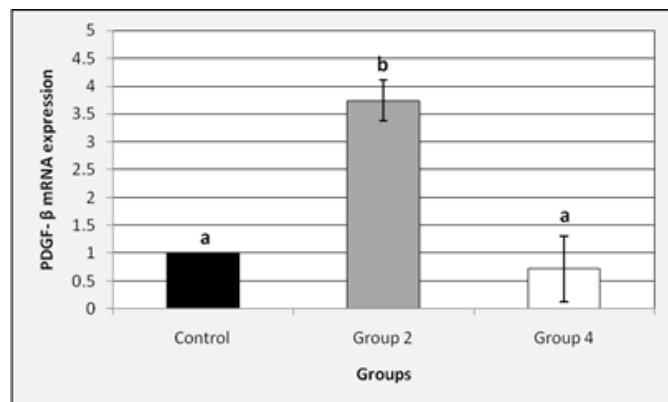


Figure 1) The study of PDGF-β mRNA expression in BAECs by Real-Time PCR.

There was a significant increase in 20 μM H₂O₂ treated cells (group 2) compared with control and 20 μM H₂O₂- 10 μM Curcumin treated cells (group 4) (p<0.05). PDGF-β: Platelet-Derived Growth Factor β; BAECs: Bovine aortic endothelial cells; Cur: Curcumin. Significant differences at p<0.05 are shown by different letters in each column.

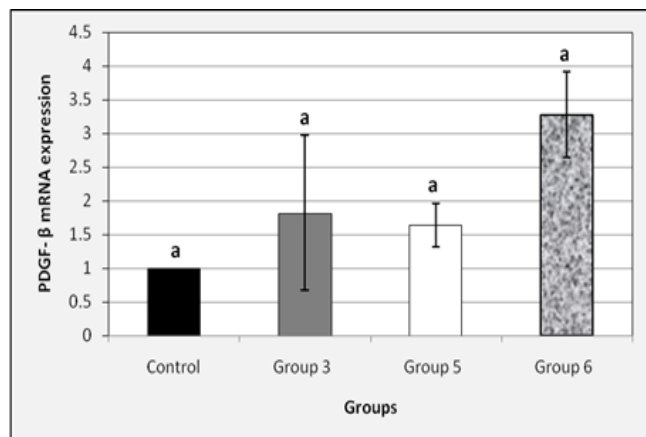


Figure 2) The levels of PDGF- β mRNA expression in BAECs were evaluated using Real-Time PCR

Group 3 (40 μM H_2O_2 -10 μM Cur/24h) showed lower levels of PDGF- β mRNA expression compared to 40 μM H_2O_2 incubated cells for 24h (group 2). Also, group 6 which received 80 μM H_2O_2 - 10 μM Cur treatment for 24h, demonstrated the highest level of PDGF- β mRNA between all experimental groups. PDGF- β : Platelet- Derived Growth Factor β ; BAECs: Bovine aortic endothelial cells; Cur: Curcumin. Significant differences at $p < 0.05$ are shown by different letters in each column.

Discussion

Atherosclerosis as a major cause of cardiovascular diseases has several risk factors such as high cholesterol and VSMCs migration (6,12). Also, the side effects of high fat diet and high cholesterol can alleviate by Curcumin (4-6). Curcumin significantly decreases fatty liver development, weight gain, dyslipidemia in Ldlr (-/-) mouse model of human atherosclerosis (4-6). This potent antioxidant modulates atherogenesis by inhibiting expression of aP_2 and CD36 in macrophages (4). Furthermore, Curcumin significantly decreases hepatic complement factor D (Cfd) and systemic CRP levels as main markers of immune complement pathway activation (5). Linton (2003) and Hassan (2011) showed that ApoE-deficient mice null for macrophage aP_2 expression,

develop significantly less atherosclerosis than controls wild type for macrophage aP_2 expression (6,13). Recent researches demonstrated strong anti-atherogenic effect of long-term Curcumin treatment in high-cholesterol treated mice. There are multiple mechanisms by which Curcumin induced anti-atherogenic effect such as alteration on lipid concentration, cholesterol concentration and immune gene expression (5). Long-term Curcumin administration has more effective and protective effect on high cholesterol-induced atherosclerosis compares to lovastatin. Most of the anti-atherogenic effects of Curcumin are similar to lovastatin. Curcumin improves lipid infiltration, ICAM-1 and VCAM-1 localization (5) and early atherosclerotic lesions (5,6). In addition, long-term Curcumin decreases plasma cholesterol, triglycerides, Apo B levels, LDL cholesterol and increases plasma HDL cholesterol and liver Apo A-I expression (5). H_2O_2 is one of the major ROS that has important roles on pathologic conditions (14). The shift in balance between oxidant (such as ROS) and antioxidant in favor of oxidants is termed "oxidative stress" (14). Many diseases including cancer (15-18), cardiovascular diseases (18-20), diabetes (20,21) and hypertension (14,22) are induced by oxidative stress (15). So, atherosclerosis can induce by ROS-related oxidative stress (23). Here, we showed that H_2O_2 as a potent oxidant can promote atherosclerosis by changes on atherosclerosis-induced genes including PDGF. H_2O_2 increased the level of PDGF- β expression in endothelial cells. Also, PDGF- β can cause VSMC proliferation and migration (24). The proliferation and migration of VSMC are critical factors for promoting atherosclerosis. Curcumin co-treatment in H_2O_2 incubated cells showed significant decrease on expression of PDGF- β mRNA versus H_2O_2 treated cells. Curcumin reduces the risk of VSMC proliferation and atherosclerosis on oxidative stress-exposed endothelial cells. ROS have been implicated in the pathogenesis of

cardiovascular diseases, in part by promoting VSMC proliferation (25-27).

Conclusion

In this study, we tested the hypothesis that Curcumin as a potent antioxidant decreases the risk of atherosclerosis by reducing H₂O₂-related oxidative stress and PDGF gene expression in endothelial cells. The understanding of the pathophysiology and effective treatment of atherosclerosis can provide new perspectives for preventive and therapeutic strategies. According to these finding, herbal diet supplement such as Curcumin can help patients or person who has risk factor of cardiovascular diseases.

Footnotes

Acknowledgments:

The authors would like to thank Falavarjan Branch, Islamic Azad University and Ahvaz Jundishapur University of Medical Sciences for providing financial supporter of this research.

Conflict of Interest:

The authors declared no conflict of interest.

References

1. von der Lohe E, Coronary Heart Disease in Women: Prevention - Diagnosis – Therapy. Springer Science & Business Media; 2003. P. 125-131.
2. Jagetia GC, Aggarwal BB. “Spicing Up” of the Immune System by Curcumin. *J Clin Immunol* 2007;27(1):19–35.
3. Antoniadis C, Tousoulis D, Stefanadis C. Effect of endothelial nitric oxide synthase gene polymorphisms on oxidative stress, inflammatory status and coronary atherosclerosis: an example of transient phenotype. *J Am Coll Cardiol* 2007;49(11):12-26.
4. Hasan ST, Zingg JM, Kwan P, Noble T, Smith D, Meydani M. Curcumin modulation of high fat diet-induced atherosclerosis and steatohepatosis in LDL receptor deficient mice. *Atherosclerosis* 2014;232(1):40-51.
5. Coban D, Milenkovic D, Chanet A, Khallou-Laschet J, Sabbe L, Palagani A, et al. Dietary curcumin inhibits atherosclerosis by affecting the expression of genes involved in leukocyte adhesion and transendothelial Mol Nutr Food Res 2012;56(8):1270-81.
6. Hasan ST, Eastwood M, Zingg JM, Meydani M. Curcumin inhibits atherosclerosis by suppressing accumulation of lipids in macrophages in LDLr^{-/-} mice. *FASEB J* 2011;25(1):339.40.
7. Sharma RA, Gescher AJ, Steward WP. Curcumin: the story so far. *Eur J Cancer* 2005;41(13):1955-68.
8. Jagetia GC, Golgod Krishnamurthy R. Curcumin Stimulates the Antioxidant Mechanisms in Mouse Skin Exposed to Fractionated γ -Irradiation. *Antioxidants* 2015;4(1):25-41.
9. Rajeswari A, Curcumin protects mouse brain from oxidative stress caused by 1-methyl-4-phenyl-1,2,3,6-tetrahydro pyridine. *Eur Rev Med Pharmacol Sci* 2006;10(4):157-161.
10. Madkour NK. Protective effect of curcumin on oxidative stress and DNA fragmentation against lambda cyhalothrin-induced liver damage in rats. *J Appl Pharm Sci* 2012;2(12):76-81.
11. Moody LR, Herbst AJ, Yoo HS, Vanderloo JP, Aiken JM. Comparative prion disease gene expression profiling using the prion disease mimetic, cuprizone. *Prion* 2009;3(2):99–109.
12. Blair HC, Sepulveda J, Papachristou DJ. Nature and nurture in atherosclerosis: The roles of acylcarnitine and cell membrane-fatty acid intermediates. *Vascul Pharmacol* 2016;15:17-23.
13. Linton MF, Fazio S. Macrophages, inflammation, and atherosclerosis. *Int Journal Obes Relat Metab Disord* 2003;27(Suppl 3):35-40.
14. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative Stress and Antioxidant Defense. *World Allergy Organ J* 2012;5(1):9–19.
15. Jenner P. Oxidative stress in Parkinson’s disease. *Ann Neurol* 2003;53(Suppl 3):26-36.
16. Lyras L, Cairns NJ, Jenner A, Jenner P, Halliwell B. An assessment of oxidative damage to proteins, lipids, and DNA in brain from patients with Alzheimer’s disease. *J Neurochem* 1997;68(5):2061–2069.
17. Sayre LM, Smith MA, Perry G. Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem* 2001;8(7):721–738.
18. Dhalla NS, Temsah RM, Netticadan T. Role of oxidative stress in cardiovascular diseases. *J Hypertens* 2000;18(6):655-673.
19. Kasparova S, Brezova V, Valko M, Horecky J, Mlynarik V. Study of the oxidative stress in a rat model of chronic brain hypo perfusion. *Neurochemical Int* 2005;46(8):601–611.
20. Kerr S, Brosnan MJ, McIntyre M, Reid JL, Dominiczak AF, Hamilton CA. Superoxide anion production is increased in a model of genetic hypertension: role of the endothelium. *Hypertension* 1999;33(6):1353-1358.

21. Kukreja RC, Hess ML. The oxygen free-radical system: from equationsthrough membrane–protein interactions to cardiovascular injury and protection. *Cardiovasc Res* 1992;26(7):641-655.
22. Asami S, Manabe H, Miyake J, Tsurudome Y, Hirano T. Cigarette smoking induces an increase in oxidative DNA damage,8-hydroxydeoxyguanosine, in a central site of the human lung. *Carcinogenesis* 1997;18(9):1763-1766.
23. Satoh K, Nigro P, Bradford C. Oxidative Stress and Vascular Smooth Muscle Cell Growth:A Mechanistic Linkage by Cyclophilin. *Antioxid redox Signal* 2010;12(5):675-782.
24. Dzau VJ, Braun-Dullaeus RC, Sedding DG. Vascular Proliferation and Atherosclerosis: New Perspectives and Therapeutic Strategies. *Nat Med* 2002;8(11):1249-1256.
25. Alexander RW. Hypertension and the pathogenesis of atherosclerosis, Oxidative stress and the mediation of arterial inflammatory re-sponse: a new perspective. *Hypertension* 1995;25(2):155–161.
26. Baas AS, Berk BC. Differential activation of mitogen-activated protein kinases by H₂O₂ and O₂ in vascularsmooth muscle cells. *Circ Res* 1995;77(1):29–36.
27. Omar HA, Cherry PD, Mortelliti MP, Burke–Wolin T, Wolin MS. Inhibition of coronary artery superox-ide dismutase attenuates endothelium-dependent and independent nitrovasodilator relaxation. *Circ Res* 1991;69(3):601-608.